Australian Rotavirus Surveillance Program annual report, 2015

Susie Roczo-Farkas, Carl D Kirkwood, Julie E Bines and the Australian Rotavirus Surveillance Group

Abstract

The Australian Rotavirus Surveillance Program, together with collaborating laboratories Australiawide, reports the rotavirus genotypes responsible for the hospitalisation of children with acute gastroenteritis during the period 1 January to 31 December 2015. During the survey period, 1,383 faecal samples were referred for rotavirus G and P genotype analysis, and of these, 1,031 were confirmed as rotavirus positive. A total of 634 specimens had been collected from children under 5 years of age, while 397 were from older children and adults. Genotype analysis of samples from both children and adults revealed that G12P[8] was the dominant genotype in this reporting period, identified in 48.2% of strains nationally. Genotype G3P[8] was the second most common strain nationally, representing 22.8% of samples, followed by G2P[4] and G1P[8] (9% and 8% respectively). G3P[8] was further divided as equine-like G3P[8] (13.2% of all strains) and other wild-type G3P[8] (9.6%). This report highlights the continued predominance of G12P[8] strains as the major cause of disease in this population. Genotype distribution was distinct between jurisdictions using RotaTeq and Rotarix vaccines. Genotype G12P[8] was more common in states using RotaTeq, while equine-like G3P[8] and G2P[4] were more common in the states and territories using Rotarix. This survey highlights the dynamic change in rotavirus genotypes observed since vaccine introduction, including the emergence of a novel equine-like G3P[8] as a major strain. The prolonged dominance of G12P[8] for a 4th consecutive year further illustrates the unexpected trends in the wild type rotaviruses circulating in the Australian population since vaccine introduction. Commun Dis Intell 2016;40(4):E527-E538.

Keywords: rotavirus, gastroenteritis, genotypes, disease surveillance

Introduction

Rotaviruses belong to the Reoviridae family and are triple layered dsRNA viruses that contain a segmented genome, consisting of 11 gene segments that encode 6 structural proteins and 6 non-structural proteins.¹ Rotaviruses are the most common cause of severe diarrhoea in young children worldwide, and are estimated to cause up to 453,000 deaths annually.² The significant morbidity and mortality associated with rotavirus infection has led to the development of vaccines, such as Rotarix[®] (GlaxoSmithKline) and RotaTeq[®] (Merck). These 2 oral live attenuated rotavirus vaccines have been shown to be safe and highly effective in the prevention of severe diarrhoea due to rotavirus infection,^{3,4} leading to both vaccines being licensed in over 125 countries and included in the national vaccination schedules of 63 predominantly high and middle-income countries worldwide.5 In Australia, rotavirus vaccines have been included in the National Immunisation Program since 1 July 2007, with excellent uptake in subsequent years. RotaTeq is administered in Queensland, South Australia, Victoria and Western Australia, while Rotarix is administered in the Australian Capital Territory, New South Wales, the Northern Territory and Tasmania.⁶

In Australia prior to vaccination being introduced, rotavirus infection had accounted for up to 10,000 childhood hospitalisations for diarrhoea each year.⁷ A significant impact on the disease burden has been observed since vaccine introduction, with studies showing a substantial decline in both rotavirus coded and non-rotavirus coded hospitalisation and emergency room visits.^{6,8–12}

The Australian Rotavirus Surveillance Program has characterised the G- and P- genotypes of rotavirus strains causing severe disease in Australian children since 1997. Data from this surveillance has shown that strain diversity as well as temporal and geographic changes occur each year; providing critical baseline data.¹³ Ongoing characterisation of circulating rotavirus genotypes in the vaccine era will provide insight into whether vaccine introduction has impacted on virus epidemiology, and altered circulating strains, which could have ongoing consequences for the success of current and future vaccination programs.

This report describes the genotype distribution of rotavirus strains causing severe gastroenteritis in Australia for the period 1 January to 31 December 2015.

Methods

Rotavirus positive specimens detected by enzyme immunoassay (EIA) or latex agglutination in collaborating laboratories across Australia were collected, stored frozen and forwarded to the Australian Rotavirus Reference Centre Melbourne, together with relevant age and sex details. The laboratories contributing samples were:

- ACT Pathology, Canberra, Australian Capital Territory
- The Virology Division, South Eastern Area Laboratory Services, Prince of Wales Hospital, New South Wales
- Virology Department, The Children's Hospital at Westmead, New South Wales
- Centre for Infectious Diseases and Microbiology, Westmead, New South Wales
- The Microbiology Department, John Hunter Hospital, Newcastle, New South Wales
- The Microbiology Department, Royal Darwin Hospital, Casuarina, Northern Territory
- The Microbiology Department, Alice Springs Hospital, Alice Springs, Northern Territory
- Forensic and Scientific Services, Queensland Health, Herston, Queensland
- Microbiology division, Pathology Queensland, Herston, Queensland
- The Queensland Paediatric Infectious Diseases laboratory, Royal Children's Hospital, Brisbane, Queensland
- Queensland Health laboratory in Townsville, Queensland
- Microbiology and Infectious Diseases Laboratory, SA Pathology, Adelaide, South Australia
- The Serology Department, Royal Children's Hospital, Parkville, Victoria
- Department of Microbiology, Monash Medical Centre, Clayton, Victoria
- Division of Microbiology, PathWest LM, The Queen Elizabeth Medical Centre, Nedlands, Western Australia.

Viral RNA was extracted from 10% to 20% faecal extracts using the QIAamp Viral RNA mini extraction kit (Qiagen) according to the manufacturer's instructions. Rotavirus G and P genotypes were determined using an in-house hemi-nested multiplex reverse transcription polymerase chain reaction (RT-PCR) assay. The first round RT-PCR reactions were performed using the Qiagen one step RT-PCR kit, using VP7 conserved primers VP7F and VP7R, or VP4 conserved primers VP4F and VP4R. The second round genotyping PCR reactions were conducted using specific oligonucleotide primers for G types 1, 2, 3, 4, 8, 9 and 12 or P types [4], [6], [8], [9],

[10] and [11].^{14–19} The G and P genotype of each sample was assigned using agarose gel analysis of second round PCR products.

First round amplicons for VP7 were also purified for sequencing by using a Wizard SV Gel for PCR Clean-Up System (Promega), according to the manufacturer's protocol. Purified DNA together with oligonucleotide primers (VP7F/R) were sent to the Australian Genome Research Facility, Melbourne, and sequenced using an ABI PRISM BigDye Terminator Cycle Sequencing Reaction Kit (Applied Biosystems, Foster City, CA, USA) in an Applied Biosystems, Foster City, CA, USA). Sequences were edited with Sequencher v.4.10.1. The genotype assignment was accomplished using BLAST (http://blast.ncbi.nlm.nih.gov/Blast.cgi) and RotaC v2.0 (http://rotac.regatools.be).²⁰

Any samples that provided a discordant result between the initial antigen detection and genotype assay were further tested using the commercial rotavirus ELISA assay ProSpecT (Thermo Fisher, Aus.), as per manufacturer's instructions to confirm the presence of rotavirus antigen.

Results

Number of isolates

A total of 1,383 faecal specimens were collected during the period 1 January to 31 December 2015 for analysis from 15 collaborating centres across Australia. For this reporting period, no samples were received from Tasmania.

A total of 1,031 samples were confirmed as rotavirus positive. Of these, 634 had been collected from children under 5 years of age, and 397 were from older children and adults. An additional 352 specimens contained either insufficient specimen for genotyping (n=7), were duplicates of samples already analysed (n=77) or the specimen was not confirmed to be positive for rotavirus (n=268) and were thus not analysed further.

Age distribution

During the 2015 reporting period, 61.5% of samples were obtained from children under 5 years of age (Table 1). Of these, one-third of all samples (33.3%) were identified in children aged 13–24 months, while the next most common age group was 25–36 months (19.7%).

Genotype distribution

All of the 1,031 confirmed rotavirus samples collected from children and adults underwent

 Table 1: Age distribution of gastroenteritis cases

Age range (months)	Age range (years)	Number	% of total	% under 5 years
0-6		120	11.6	18.9
7–12	≤1	82	8.0	12.9
13–24	1–2	211	20.5	33.3
25–36	2–3	125	12.1	19.7
37–48	3–4	53	5.1	8.4
49-60	4–5	43	4.2	6.8
Sub-total		634	61.5	_
61–120	5–10	139	13.5	
121–240	10–20	48	4.7	
241–960	20–80	165	16.0	
961+	>80	45	4.4	
Total		1,031	_	

genotype analysis (Table 2). G12P[8] strains were the most common genotype identified nationally, representing 48.2% of all specimens analysed. This genotype was identified as the dominant type in 4 states, Queensland, Victoria, Western Australia and South Australia, representing 53.8%, 80.2%, 65.4% and 50.8% of strains respectively.

G3P[8] strains were the second most common genotype identified nationally, representing 22.8% of all specimens. These were further divided as equine-like G3P[8] (13.2%) and other wild-type G3P[8] (9.6%). Equine-like G3P[8] were identified in all 7 states or territories that submitted samples, and was the dominant type in the Australian Capital Territory and New South Wales, where it represented 96% and 67.2% respectively. In the Northern Territory, equine-like G3P[8] was the second most common type after G2P[4], representing 36.4% and 48.6% of strains respectively. No other wild-type G3P[8] strains were detected in Rotarix states.

G2P[4] and G1P[8] strains were the third and fourth most common genotypes nationally, representing 8.6% and 8.1% of all specimens respectively. G1P[8] was identified in all locations except the Australian Capital Territory, and was the third most common strain in Queensland and South Australia, representing 11.3% and 13.2% of strains respectively.

Thirty samples (3% of all strains) were categorised into 9 uncommon G- and P- genotype combinations (Table 2). Six G3P[3], 5 G8P[14], 4 G9P[4], and 4 G12P[6] strains were identified. Three G1P[4] strains were also noted, as were 3 G8P[8] strains. A further 2 G8P[14], 2 G12P[4] and a single G10P[14] strain were identified. A G- or P- genotype could not be assigned to 35 samples, of which 23 (65.7%) were observed in Western Australia. Of the 35 samples, 31 were G- non-typeable, 2 were P- non-typeable and 2 had no G- or P- genotype assigned, although they were positive by EIA. The partially non-typeable samples could be strains that contain unusual or uncommon G- or P- genotypes and would not be typeable with the primers used. While this can also be suggested for the EIA positive/G- and Pnon-typeable samples, another possible explanation is that the extracted RNA for these samples contained inhibitors that could have prevented the function of the enzymes used in the RT and/or PCR steps.

Thirty-eight samples were identified that contained a strain that was a component of the RotaTeq or Rotarix vaccine. Such Rotarix strains were found in New South Wales (n=1), and the Northern Territory (n=2), while RotaTeq strains were identified in South Australia (n=5), Victoria (n=1) and Western Australia (n=29). In each instance, a vaccine component was determined by RT-PCR and confirmed by sequence analysis of the VP6 and VP7 gene.

Genotypes identified in samples from children less than 5 years of age

In total, 634 rotavirus samples were collected from children under 5 years of age (Table 3). In this cohort, genotype G12P[8] strains were the most commonly identified, found in 41.6% of samples. G3P[8] was the second most common genotype (23.5%), comprising of 14.8% equine-like G3P[8] and 8.7% other wild-type G3P[8] (Figure 1). G2P[4] strains were the third most common genotype (10.1%). G1P[8] and G9P[8] strains represented minor genotypes in this cohort, and were identified in 8.8% and 2.2% of samples respectively (Table 3).

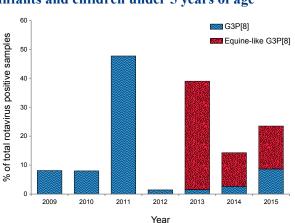


Figure 1: Wild-type G3P[8] occurrences in infants and children under 5 years of age

TADIC 2: INDIATILUS O ANU T ZUTOLY DE UISU IDUUTOTI IN INTANUS, ENTRUCH ANU AUUTIS, I VAIUALY TO ST DECEMBEL 2013		-9 T NT	I CANTON I	mann of	ommon		- 600														
	Tvne	G1P[8]	8	G2P[4]	4	G3P[8]		G3P[8]*		G4P[8]	G9P[8]	[8]	G12P[8]	[8]	Non-type [†]		Vaccine [‡]	++	Other	Neg	Insuff
Centre	total	%	c	%	c	u %	%	L	%	c	%	c	%	c	%	c	u %	%	u v	2	2
Australian Capital Territory	Territo	٢																			
ACT	25	I	0	I	0	1	0 96	3 24	1	0	I	0	I	0	I	0	1	0	4	-	0
New South Wales																					
POW	0	I	0	I	0	1	0	0	1	0	Ι	0	I	0	I	0	1	0	0	0	0
Westmead	35	ი	~	6	e	1	0 71	1 25	1	0	ო	-	9	7	ი	-	1	0	6 2	7	0
John Hunter	20	10	7	I	0	-	0 60) 12	I	0	15	з	5	-	I	0	5	-	5 1	9	0
NSW subtotal:	55	5	ო	വ	с	1	0 67	, 37	1	0	2	4	5	ო	7	-	N	-	5 3	8	0
Northern Territory																					
Alice Springs	15	I	0	93	14	1	0	0	1	0	I	0	I	0	I	0	1	0	7 1	7	0
Darwin	48	I	0	35	17	1	0 54	1 26		0	10	2	I	0	I	0	1	0	0	ო	0
Western Diagnostic	12	I	0	I	0	1	0 100) 12	1	0	1	0	I	0	I	0	1	0	0	0	0
Other [§]	31	3	-	68	21	3 1		0	I	0	I	0	I	0	6	2	6	2 13	3 4	10	2
Northern Territory subtotal:	106	-	-	49	52	<i>د</i>	1 36	38	1	0	വ	ъ	I	0	5	7	2	N	ນ	20	2
Queensland																					
Pathology Brisbane	61	10	9	11	7	7	4 15	6	1	0	വ	ო	46	28	ო	2	1	0	3	თ	0
Qld Regional	0	I	0	I	0	1	۱ 0	0	1	0	I	0	I	0	Ι	0	1	0	0	ε	0
Pathology Townsville	19	16	ო	I	0	1	і о	0	1	0	ъ	-	79	15	I	0	1	і 0	0	ო	0
Queensland subtotal:	80	7	ര	ი	~	5	4	6	1	0	വ	4	54	43	ო	7	1	0	3	15	0
South Australia																					
Adelaide	418	13	55	5	22	21 89	3	3 12	$\overline{\mathbf{v}}$	-	۲	-	51	212	7	7	1	2	3 14	100	2
Victoria														-							
RCH	80	9	5	I	0	1	6 0	7	1	0	-	-	80	64	I	0		~	3 2	30	0
Monash	-	I	0	I	0	1	ו 0	0	1	0	1	0	100	-	I	0	1	 0	0	0	0
Victoria subtotal:	81	9	5	I	0	1	6	6	1	0	-	-	80	65	I	0	.		2	30	0

2 5 2 5 3 9 <u>–</u> 89 10 99 13 136 <1
Equine-like ଓ୬୮/୪୦ A specimen where G and/or P genotype was not determined.
A specimen where a vaccine component from RotaTeq or Rotarix was found.
Faecal specimens from the Northern Territory which were processed in Adelaide.
4x G-nt P[8]; 1x G-nt P[6]; 1x G4P[nt] and 1x nt (EIA positive)

Table 2 continued: Rotavirus G and P genotype distribution in infants, children and adults, 1 January to 31 December 2015

Australian Rotavirus Surveillance Program, 2015

TADIE J. INUAVILUS U AIRU I GERULY DE UISULIDUUUR III IRITARIES		IULYP	ngin a	Innat						anu chiniten unuel o years ut age, i januar y tu ol december 2010		age, 1	Jallua	. n v			C107					
	Type	G1P[8]	[8]	G2P[4]	4	G3P[8]		G3P[8]*		G4P[8]	د ق	G9P[8]	G13	G12P[8]	Non-type [†]	/pe [†]	Vaccine [‡]		Other	Neg	Ë	ıff
Centre Australian Canital Territory	IOI	<u></u>	-	<u>«</u>	-	%				= %	<u>ج</u>	-	۶	-	<u>ج</u>	-	<u>¢</u>	=	= %	-		
	14	1	c		c		с С	03	13			c	_	c	1	c		c	7	_	c	
New South Wales	:		>				-		-		_	>				>				-	-	
POW	0	ī	0	ı	0	1	0		_	0	1	0	1	0	ı	0	1	0	0	-	-	Ľ
Westmead	23	4	~	4	-	I		~	. 18	0	4	~	I	0	4	~	I	0	4	_	0	
John Hunter	10	I	0	I	0	I	0	60	9	0	20	2	I	0	I	0	10	-	10 1	-		
NSW subtotal:	33	ო	-	ო	-	I	0	73 2	24	0	တ	ო	I	0	ო	-	ო	-	6 2	-	0	
Northern Territory	-				-				-		-			-		-				-		
Alice Springs	14	ī	0	93	13	I	0	1	0	0	I	0	I	0	Т	0	I	0	7 1			
Darwin	44	I	0	34	15	I	0	55 2	24	0	7	5	I	0	I	0	I	•	0	1	1	
Western Diagnostic	12	I	0	I	0	I	0	100 1	12	0		0	I	0	I	0	I		0	0	0	
Other ^s	27	4	-	64	18	I	0	I		0	1	0	I	0	7	2	7	2	15 4			
Northern Territory subtotal:	97	-	-	47	46	I	0	37 3	36	0	5	Ω	I	0	7	2	7	5	5 5	17	2	
Queensland																						
Pathology Brisbane	24	17	4	4	-	4		17		0		0	54	13	4	-	I		1			
Qld Regional	0	I	0	I	0	I	0	I		0		0	I	0	I	0	I	•	0	ю —		
Pathology Townsville	80	13	-	I	0	I	0	I	0	-	13	-	75	9	I	0	I		1		0	
Queensland subtotal:	32	16	ъ	ო	-	ო	-	13	4	0	ۍ ا	-	59	19	ო	-	I	•	0	12	0	
South Australia																						
Adelaide	238	15	36	9	15	22	52	ო	v ∞	-1	V	-	45	107	7	4	7	5	4	39	ო	
Victoria																						
RCH	55	7	4	I	0	I	0	7	4	0	7	-	78	43	I	0	7	~	4 2	17		
Monash	-	Т	0	Т	0	I	0			0	۱ 	0	100	-	Т	0	I	0	1	0	0	
Victoria subtotal:	56	7	4	I	0	I	0	7	4	0	2	-	79	44	0	0	7	1	4 2	17	0	
Western Australia																						
PathWest	164	5	6	-	-	٢	2	Э	2	0	7	З	57	94	12	19	18	29	1 2	55	0	
Total	634	6	56	10	64	റ	55	15 5	94 <	1	2	4	42	264	4	27	9	38	3 21	142	ນ	
											Č	.solames routes										
t Δ snariman whara G and/or D ranotyna was not datarminad	nd/or D c	henotyr		not dete	rminer	_						Australian Canital Territory 1v G12P[6]	eo. anital Te	rritory 1	ч G12D	[9]						
 A speciment where a varcine commonent from RotaTen or Rotariy was found 			on from	RotaTe		otariy wa	s found				enc Mel	New South Wales	apitai is Vales		1× G 8 14] 1× G 9 14]	[4] 17 (
	the Nor	thern 7	erritory	which v		ocessed	in Adelaide.	aide.			Nor	Northern Territory	itory		1x G1P[4], 4x G3P[3]	1, 4× G	3P[3]					
eg			•								Sou	South Australia	ia ,	-	× G1P[₄	ij, 1× G	3P[3], 2)	(G8P[4]	1x G1P[4], 1x G3P[3], 2x G8P[4], 2x G9P[4], 1x G10P[14],	[4], 1× (G10P[14	÷
Insuff Insufficient sample for testing	esting													⁽ N	x G12P	4						
POW Prince of Wales Hospital	a										Victoria	oria .	:	,	1x G9P[4], 1x G12P[6]	ij, 1× G	12P[6]					
RCH Royal Children's Hospital	tal										Wes	Western Australia	tralla	-	1x G3P[3], 1x G8P[14]	ם', 1× ק	8P[14]					

Genotypes identified in samples from individuals greater than 5 years of age

A total of 397 rotavirus samples were collected from children over the age of 5 years, and adults (Table 4). In this cohort, genotype G12P[8] strains were the most common, found in 58.4% of samples. G3P[8] strains (including equine-like G3P[8]) were the second most common genotype (21.7%), and G1P[8] and G2P[4] strains were equal third most common genotypes, identified in 6.8% and 6.3% respectively. Genotype G9P[8] strains were identified in 2.3% of samples.

Distribution of genotypes according to vaccine type

G and P genotypes from the rotavirus samples collected from infants and children under 5 years of age by vaccine usage were compared (Figure 2). In states where RotaTeq is in use, G12P[8] strains were the dominant genotype in children less than 5 years of age, identified in 54% of samples, while G3P[8] were the second most common, identified in 15.5% of strains. These G3 were further divided as equine-like G3P[8] (4.3%) or other wild type G3P[8] (11.2%). G1P[8] strains were the third most common genotype representing 11% of samples. In locations using Rotarix, equine-like G3P[8] strains were dominant, identified in 50.3% of strains, followed by G2P[4], identified in 32.4% of samples.

Consistency in genotype distribution among states using RotaTeq vaccine was observed, with all 4 RotaTeq states (Queensland, Victoria, South Australia and Western Australia) having G12P[8] as the dominant genotype. However, in states and territories using Rotarix (New South Wales, the Australian Capital Territory and the Northern Territory), the dominant genotype differed, with G2P[4] dominant in Northern Territory and equine-like G3P[8] dominant in New South Wales and the Australian Capital Territory.

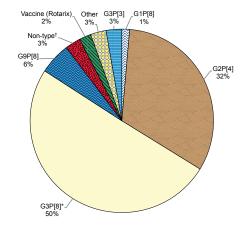
Discussion

This 2015 Australian Rotavirus Surveillance report describes the annual distribution of rotavirus genotypes and geographic differences in genotypes causing disease in Australia for the period 1 January to 31 December 2015. Genotype G12P[8] remained the dominant genotype nationally, representing 48.2% of all strains from all age groups. Genotype G3P[8] was the second most common genotype nationally, comprising 23% of all strains, but was the dominant genotype in New South Wales and the Australian Capital Territory only. Genotype G2P[4] and G1P[8] were the third and fourth most common, representing 8.6% and 8.1% of all strains respectively. In the samples collected from infants and children under 5 years of age, genotype G12P[8] continued to remain as the dominant genotype nationally, identified in 41.6% of samples. The second most common genotype was G3P[8], identified in 23.5% of samples, while the third most common was G2P[4], identified in 10% of samples.

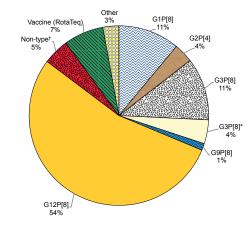
In Australia, G12P[8] emerged in 2012 and subsequently became the dominant strain in 2013 and 2014, representing 33% and 29.6% of all strains respectively.^{19,22,23} This figure increased in 2015 to encompass 48.2% of all strains, and has been identified as the dominant strain in all 4 states that use the RotaTeq vaccine (South Australia, Queensland, Victoria and Western Australia). Similar observations have been observed in

Figure 2: Overall distribution of rotavirus G and P genotypes identified in Australian children based on vaccine usage, Australia, 1 January to 31 December 2015

Rotarix states[†]







- * Equine-like G3P[8]²¹
- † The Australian Capital Territory, New South Wales, and the Northern Territory
- ‡ Queensland, South Australia, Victoria, and Western Australia

TADIE 4: ROLAVITUS & AIRU F GENOLYPE UISUTIDULION IN CHIRUTEN	u r ge	IIOUY	sin ar	Innaria		CIIII		VCI J J	Cars	ui age	allu a	uurts,	т лап	uary		Decel	over 5 years of age and addits, I January to 51 December 201	CT					
	Tvne	G1P[4]	P[4]	G1P[8]		G2P[4]		G3P[8]		G3P[8]*	G8P[8]	[8]	G8P[14]	14]	G9P[8]		G12P[6]	<u>G</u>	G12P[8]	Non-type [†]		Neg I	Insuff
Centre	total	%	c	%	c	%	c	n %	%	c	%	c	%	c	%	°` c	u %	%	c	%	c	c	c
Australian Capital Territory	>																						
ACT	11	I	0	I	0	I	0	1	0 100	11	1	0	I	0	I	- 0	0	1	0	I	0	0	0
New South Wales																							
POW	0	Ι	0	I	0	I	0	1	۱ 0	0	I	0	I	0	I	0	0	I	0	I	0	0	0
Westmead	12	I	0	I	0	17	N	1	0 58	8	œ	-	I	0	I	0	0	17	2	I	0	2	0
John Hunter	10	I	0	20	2	I	0	-	0 60	9 (I	0	I	0	10	-	- 0	10	1	I	0	5	0
NSW subtotal	22	I	0	6	7	6	~		0 59	13	2	-	I	0	4	- -	0	4	с	I	0		0
Northern Territory																							
Alice Springs	-	I	0	I	0	100	~	-	 0	0	1	0	I	0	I	0	0		0	I	0	0	0
Darwin	4	I	0	I	0	50	2		0 50)	I	0	I	0	I	- 0	0	1	0	I	0	0	0
Western Diagnostic	0	I	0	I	0	I	0		0	0	I	0	I	0	I	0	0	I	0	I	0	0	0
Other [‡]	4	I	0	I	0	75	3	25 .	-	0	I	0	I	0	I	0	- 0	I	0	I	0	З	0
Northern Territory subtotal	ი	I	0	I	0	67	۰ و	11	1 22	5	I	0	Т	0	I	- 0	0	1	0	I	0	e	0
Queensland																							
Pathology Brisbane	37	I	0	5	2	16	9	∞	3 14	1	ო	-	ო	~	œ	ო	0	4	15	ო	-	2	0
Qld regional	0	I	0	I	0	I	0	-	ו 0	0	I	0	I	0	I	0	0	I	0	I	0	0	0
Pathology Townsville	11	I	0	18	2	Т	0		ו 0	0	1	0	Т	0	I	- 0	0	73	ω	6	-	-	0
Queensland subtotal:	48	I	0	ω	4	13	9	9	3 10	5	2	-	7	-	9	ო	0	48	23	4	2	e	0
South Australia																							
Adelaide	180	-	-	11	19	4	~	21 37	7 2	4	1	0	-	2	I	0	1 2	58	105	7	e	61	2
Victoria																						:	
RCH	25	I	0	4	~	I	0	1	0 12	3	I	0	I	0	I		0	84	21	I	0	13	0
Monash	0	I	0	I	0	Т	0	1	। ०	0	I	0	I	0	Т	' 0	0	1	0	I	0	0	0
Victoria subtotal:	25	Ι	0	4	٦	I	0		0 12	3	1	0	I	0	I	0	0	84	21	I	0	13	0
Western Australia																							
PathWest	102	I	0	۲	-	4	4	с С	3 4	4	-	-	I	0	5	۔ 2	0	78	80	4	4	37	0
Total	397	v	-	4	27	9	25	11 44	4	42	~	ო	-	ო	7	້ ດ	1 2	58	232	7	ი	124	2
* Eauine-like G3P[8]											Nea	Neo	Negative for rotavirus	or rotav	irus								
A specimen where G and/or P genotype was not determined.	and/or	P gen	otype v	/as not d	etermi	ned.					finsul		Insufficient sample for testing	sample	e for te	sting							
Faecal specimens from the Northern Territory which were processed in Adelaide.	om the h	Vorthe	'n Terri	tory whic	h wer	e proce	ssed ir	λ Adelaic	Je.		POW		Prince of Wales Hospita	Vales F.	lospital								

RCH Royal Children's Hospital.

Nicaragua, where RotaTeq was added to the infant schedule in 2006; more than a 2-fold increase in rotavirus cases in children under 5 years of age was attributed to a G12 outbreak in the 2012/2013 season.²⁴ Since 2000, over 30 countries have reported an emergence or re-emergence of G12 rotavirus strains.^{25–27} Furthermore, G12P[8] have been identified as the dominant strain in multiple countries since 2010.24,28-31 During the 2010-2011 season, 65% of all strains in Basque Country, Spain were identified as G12P[8].³¹ At this time, vaccine coverage was estimated to be below 30%, even though both vaccines were available in Basque Country since 2006–2007.³¹ In 2013, G12P[8] represented 82% of all strains in Nicaragua; 86% in St Louis, Missouri and 89% in Atlanta, Georgia.24,29,30 In addition, G12P[8] emerged as a dominant strain in Brazil, from 22.7%–27.3% in 2012 to 86.6% in 2014, although G12 had decreased to represent only 2.7% of all strains in 2013.^{28,32,33}

Whilst not yet dominant, G12 has been identified as an important genotype in countries such as Italy, Saudi Arabia, Cameroon, and North and South America.^{34–38} In Delhi, India, a study investigating strain diversity of samples collected from 2007–2012, established that G12 represented 14.8% of all genotypes identified. The majority of G12 strains found in the study had VP4 genotype P[6] (67.4%), while P[8] represented only 7% of all G12 cases. Another combination, G12P[4], was identified and represented 14% of all G12 strains identified.³⁹ G12P[4] and G12P[6] have also been identified in Australia in 2013 and 2015, however they were uncommon and represented less than 1% for each surveillance period. Taken together, these data demonstrate that G12 strains are an important cause of rotavirus disease both in Australian and global settings.

This survey reports the persistence of G3P[8] strains as the second most common genotype across Australia. Previously, G3P[8] strains generally represented 4% to 11% of strains over a 1-year reporting period. However, in 2013, G3P[8] emerged as the second most common genotype, representing 31% of all strains.¹⁹ Full genome analysis of these G3P[8] samples revealed that in 2013, a novel inter-genogroup reassortant strain (denoted as equine-like G3P[8], or G3P[8]* in Figure 2) had emerged, which had a distinct VP7 antigenic profile compared to other wild-type Australian G3P[8] strains.²¹ The genome constellation of equine-like G3P[8] contained an equine-like G3 VP7, a P[8] VP4 and a genogroup 2 backbone I2-R2-C2-M2-A2-N2-T2-E2-H2.21 This strain continued to persist, representing 37% of all strains in 2013, 12% in 2014 and 15% of 2015 samples from infants and children under 5 years of age (Figure 1).

The state and territory based tender process has resulted in the use of either RotaTeq or Rotarix vaccines in states and territories in Australia and provides a unique opportunity to observe and compare rotavirus strain diversity. Since vaccine introduction, significant differences in genotype distribution in infants and children under 5 years of age have been observed between jurisdictions using each vaccine.⁴⁰ In the current survey, equine-like G3P[8] was dominant in locations using Rotarix (50.3%), although no other wildtype G3P[8] strains were identified; whereas in RotaTeq states, other wild-type G3P[8] were more common (11.25%) compared with equine-like G3P[8] (4.3%).

The emergence of G12P[8] and novel genotypes globally have raised concerns regarding vaccine efficacy; however a review of vaccine trials and surveillance studies up to 2012 demonstrated that in high income countries such as Belgium and the United States of America, the monovalent Rotarix (G1P[8]) and pentavalent RotaTeq (G1-4, P[8]) vaccines continue to be highly efficacious against commonly circulating rotavirus strains.⁴¹ The pooled effectiveness of Rotarix was 94% against homotypic strains, to 87% against fully heterotypic strains.⁴¹ RotaTeq exhibited similar effectiveness; from 83% against homotypic strains to 75% against singleantigen non-vaccine type strains.⁴¹ Nevertheless, a recent study from Belgium reported that multiple deduced amino acid differences existed between the VP7 and VP4 antigenic epitopes of vaccine and currently circulating strains, such as G12 and G9.42 The VP7 proteins of G12P[8] strains alone contained 9 amino acid differences compared with the strains of RotaTeq, and 16 amino acid differences to Rotarix strain.⁴² These differences, together with the high evolutionary rate of the G12 VP7 gene (1.66 x 10^{-3} substitutions/site/year), raise the concern that this strain could ultimately escape the rotavirus neutralising antibody response induced by vaccines.^{27,42} Vaccine effectiveness studies are required to determine if current vaccines are able to protect successfully against novel strains, such as the equine-like G3P[8] and G12P[8].

The continued dominance of G12P[8] over a 4-year period in states using RotaTeq support the concept hypothesis that vaccine-related selective pressure may be arising. Vaccine-related selective pressure have also been implicated in the persistence of G2P[4] in countries such as Belgium, where Rotarix provides high vaccine coverage.^{43,44} This observation is of particular interest, as the 2015 Australian data described in this report found that in infants and children under 5 years of age, there was a marked increase in G2P[4] cases to 32% in jurisdictions using Rotarix, reflecting a trend also observed in 2012.⁴⁵ In this reporting period, over 1,000 rotavirus positive samples were collected from across Australia. Caution in interpreting the data is recommended, as the increase in the number of samples received may not necessarily reflect the number of cases being greater, since not all samples are forwarded by collaborating hospitals and diagnostic laboratories. Regardless, a robust increase in the number of samples sent from South Australia, Western Australia and Victoria have been noted, particularly in 2013 and 2015 in which the majority of samples from these states were genotyped as G12P[8]. The emergence of G12 and novel strains such as the equine-like G3P[8], G3P[3], G10P[14] and G8 strains demonstrate a dynamic virus population in the current post vaccine era. Therefore, the continued variations in the wild type strain population will remain a challenge to vaccine effectiveness and will require continued monitoring.

Acknowledgements

The Rotavirus Surveillance Program is supported by grants from the Australian Government Department of Health, GlaxoSmithKline and CSL. Dr CD Kirkwood is supported by a NHMRC senior research fellowship. The Murdoch Childrens Research Institute (MCRI) is supported by the Victorian Government's Operational Infrastructure Support program.

We thank H Tran and S Thomas for providing technical assistance.

Rotavirus positive specimens were collected from numerous centres throughout Australia. The significant time and effort involved in the collection, storage, packaging, compiling data and forwarding of specimens was much appreciated.

The National Rotavirus Surveillance Group includes:

Australian Rotavirus Reference Centre

Mrs Susie Roczo-Farkas; Coordinator, Research Assistant, Murdoch Children's Research Institute, Victoria

Associate Prof Carl Kirkwood; Director (prior to August 2015), Enteric Virus Group, MCRI

Prof Julie Bines; Director (after August 2015), Enteric Virus Group, MCRI

Australian Capital Territory

Mr C Moffat, Ms E Malinsky and members of ACT Pathology, Canberra Hospital

New South Wales

Professor W Rawlinson, Mr J Merif and members of the Virology Division, SEALS, Prince of Wales Hospital

Dr A Kesson, Ms I Tam and members of the Virology Department, The Children's Hospital at Westmead

Dr V Sintchenko, T Olna, Centre for Infectious Diseases and Microbiology, Westmead Hospital

Dr R Givney, S Pearce, K Delves and members of the Microbiology Department, John Hunter Hospital, Newcastle

Northern Territory

Dr R Baird, Ms J Hennessy, Ms P Smith and members of the Microbiology Department, Royal Darwin Hospital, Tennant Creek Hospital, Gove District Hospital and Katherine District Hospital

Mr J McLeod and members of the Microbiology Department, Alice Springs Hospital, Alice Springs

Ms H Cook, Centre for Disease Control, Darwin

Queensland

Mr F Moore, Ms J McMahon, Forensic and Scientific Services, Queensland Health, Herston

Dr G Nimmo, Dr M Nissen, Ms S Ye and department members, Microbiology division, Pathology Queensland Central laboratory, Herston

Dr S Lambert, Ms C Bletchly and members of the Queensland Paediatric Infectious Diseases laboratory, Royal Children's Hospital, Brisbane

Ms G Gilmore and members of the Queensland Health Laboratory, Townsville

South Australia

Professor G Higgins, Ms S Schepetiuk and members of the Microbiology and Infectious Diseases Laboratory, SA Pathology, Adelaide

Victoria

Miss P Adamopolous and members of the Serology Department, Royal Children's Hospital, Parkville

Dr J Buttery, Mrs D Kotsanas and members of the Department of Microbiology, Monash Medical Centre, Clayton

Western Australia

Dr D Smith, Dr A Levy, Ms J Wuillemin and members of Division of Microbiology, PathWest LM, Nedlands

Author details

Mrs Susie Roczo-Farkas,¹ Research Assistant, Murdoch Children's Research Institute

 $\mathsf{Dr}\ \mathsf{Carl}\ \mathsf{D}\ \mathsf{Kirkwood},^2$ Senior Program Officer, Bill and Melinda Gates Foundation

Prof Julie E Bines,^{1,3,4} Group Leader, Enteric Virus Group and Rotavirus Group, Murdoch Children's Research Institute and the Australian Rotavirus Surveillance Group

- 1. Enteric Virus Group, Murdoch Childrens Research Institute, Parkville, Victoria
- 2. Bill and Melinda Gates Foundation, Seattle, USA
- 3. Department of Paediatrics, The University of Melbourne, Victoria
- 4. Department of Gastroenterology and Clinical Nutrition, Royal Childrens Hospital, Parkville, Victoria

Corresponding Author: Mrs Susie Roczo-Farkas, Enteric Virus Group, Level 5, Murdoch Childrens Research Institute, Royal Children's Hospital, Flemington Road, Parkville, Victoria, 3052. Telephone: +61 3 8341 6383. Email: susie.roczofarkas@mcri.edu.au

References

- Estes M, Kapikian A. Rotaviruses. In: Fields virology. 5th edn. Philadelphia: Wolters Kluwer Health/Lippincott Williams and Wilkins; 2007. p. 1917–1974.
- Tate JE, Burton AH, Boschi-Pinto C, Steele AD, Duque J, Parashar UD. 2008 estimate of worldwide rotavirusassociated mortality in children younger than 5 years before the introduction of universal rotavirus vaccination programmes: a systematic review and meta-analysis. Lancet Infect Dis 2012;12(2):136–141.
- Vesikari T, Matson DO, Dennehy P, Van Damme P, Santosham M, Rodriguez Z, et al. Safety and efficacy of a pentavalent human-bovine (WC3) reassortant rotavirus vaccine. N Engl J Med 2006;354(1):23–33.
- Ruiz-Palacios GM, Perez-Schael I, Velazquez FR, Abate H, Breuer T, Clemens SC, et al. Safety and efficacy of an attenuated vaccine against severe rotavirus gastroenteritis. N Engl J Med 2006;354(1):11–22.
- 5. PATH. Rotavirus vaccine access and delivery. Available from: http://sites.path.org/rotavirusvaccine/
- Buttery JP, Lambert SB, Grimwood K, Nissen MD, Field EJ, Macartney KK, et al. Reduction in rotavirus-associated acute gastroenteritis following introduction of rotavirus vaccine into Australia's National Childhood vaccine schedule. Pediatr Infect Dis J 2011;30(1 Suppl):S25–S29.

- Carlin JB, Chondros P, Masendycz P, Bugg H, Bishop RF, Barnes GL. Rotavirus infection and rates of hospitalisation for acute gastroenteritis in young children in Australia, 1993–1996. Med J Aust 1998;169(5):252–256.
- Dey A, Wang H, Menzies R, Macartney K. Changes in hospitalisations for acute gastroenteritis in Australia after the national rotavirus vaccination program. *Med J Aust* 2012;197(8):453–457.
- Lambert SB, Faux CE, Hall L, Birrell FA, Peterson KV, Selvey CE, et al. Early evidence for direct and indirect effects of the infant rotavirus vaccine program in Queensland. *Med J Aust* 2009;191(3):157–160.
- Pendleton A, Galic M, Clarke C, Ng SP, Ledesma E, Ramakrishnan G, et al. Impact of rotavirus vaccination in Australian children below 5 years of age: a database study. *Hum Vaccin Immunother* 2013;9(8):1617–1625.
- Davey HM, Muscatello DJ, Wood JG, Snelling TL, Ferson MJ, Macartney KK. Impact of high coverage of monovalent human rotavirus vaccine on Emergency Department presentations for rotavirus gastroenteritis. Vaccine 2015;33(14):1726–1730.
- David RL, Kirk MD. Rotavirus gastroenteritis hospitalisations following introduction of vaccination, Canberra. Commun Dis Intell 2014;38(1):E3–E8.
- Kirkwood CD, Boniface K, Bogdanovic-Sakran N, Masendycz P, Barnes GL, Bishop RF. Rotavirus strain surveillance—an Australian perspective of strains causing disease in hospitalised children from 1997 to 2007. Vaccine 2009;27 Suppl 5:F102–F107.
- Gouvea V, Glass RI, Woods P, Taniguchi K, Clark HF, Forrester B, et al. Polymerase chain reaction amplification and typing of rotavirus nucleic acid from stool specimens. J Clin Microbiol 1990;28(2):276–282.
- Gentsch JR, Glass RI, Woods P, Gouvea V, Gorziglia M, Flores J, et al. Identification of group A rotavirus gene 4 types by polymerase chain reaction. J Clin Microbiol 1992;30(6):1365–1373.
- Simmonds MK, Armah G, Asmah R, Banerjee I, Damanka S, Esona M, et al. New oligonucleotide primers for P-typing of rotavirus strains: Strategies for typing previously untypeable strains. J Clin Virol 2008;42(4):368–373.
- Banerjee I, Ramani S, Primrose B, Iturriza-Gomara M, Gray JJ, Brown DW, et al. Modification of rotavirus multiplex RT-PCR for the detection of G12 strains based on characterization of emerging G12 rotavirus strains from South India. J Med Virol 2007;79(9):1413–1421.
- Itturiza-Gomara M, Cubitt D, Desselberger U, Gray J. Amino acid substitution within the VP7 protein of G2 rotavirus strains associated with failure to serotype. J Clin Microbiol 2001;39(10):3796–3798.
- Kirkwood CD, Roczo-Farkas S, Australian Rotavirus Surveillance Group. Australian Rotavirus Surveillance Program annual report, 2013. Commun Dis Intell 2014;38(4):E334–E342.
- Maes P, Matthijnssens J, Rahman M, Van Ranst M. RotaC: a web-based tool for the complete genome classification of group A rotaviruses. BMC Microbiol 2009;9:238.
- Cowley D, Donato CM, Roczo-Farkas S, Kirkwood CD. Emergence of a novel equine-like G3P[8] intergenogroup reassortant rotavirus strain associated with gastroenteritis in Australian children. J Gen Virol 2016;97(2):403–410.

- 22. Kirkwood CD, Roczo-Farkas S, Bishop RF, Barnes GL, Australian Rotavirus Surveillance Group. Australian Rotavirus Surveillance Program annual report, 2012. Commun Dis Intell 2014;38(1):E29–E35.
- Kirkwood CD, Roczo-Farkas S, Australian Rotavirus Surveillance Group. Australian Rotavirus Surveillance Program annual report, 2014. Commun Dis Intell 2015;39(3):E337–E346.
- 24. Bucardo F, Mercado J, Reyes Y, Gonzalez F, Balmaseda A, Nordgren J. Large increase of rotavirus diarrhoea in the hospital setting associated with emergence of G12 genotype in a highly vaccinated population in Nicaragua. *Clin Microbiol Infect* 2015;21(6):603, e601–e607.
- 25. Banyai K, Laszlo B, Duque J, Steele AD, Nelson EA, Gentsch JR, et al. Systematic review of regional and temporal trends in global rotavirus strain diversity in the pre rotavirus vaccine era: insights for understanding the impact of rotavirus vaccination programs. *Vaccine* 2012;30 Suppl 1:A122–A130.
- Iturriza-Gomara M, Dallman T, Banyai K, Bottiger B, Buesa J, Diedrich S, et al. Rotavirus genotypes cocirculating in Europe between 2006 and 2009 as determined by EuroRotaNet, a pan-European collaborative strain surveillance network. *Epidemiol Infect* 2011;139(6):895–909.
- Matthijnssens J, Heylen E, Zeller M, Rahman M, Lemey P, Van Ranst M. Phylodynamic analyses of rotavirus genotypes G9 and G12 underscore their potential for swift global spread. *Mol Biol Evol* 2010;27(10):2431–2436.
- 28. Luchs A, Cilli A, Morillo SG, de Souza Gregorio D, de Souza KA, Vieira HR, et al. Detection of the emerging rotavirus G12P[8] genotype at high frequency in Brazil in 2014: Successive replacement of predominant strains after vaccine introduction. Acta Trop 2016;156:87–94.
- 29. Immergluck LC, Parker TC, Jain S, Laghaie E, Spandorfer P, Jerris RC, et al. Sustained effectiveness of monovalent and pentavalent rotavirus vaccines in children. J Pediatr 2016;172:116–120.
- Wylie KM, Weinstock GM, Storch GA. Emergence of rotavirus G12P[8] in St. Louis during the 2012–2013 rotavirus season. J Pediatric Infect Dis Soc 2015;4(4):e84– e89.
- Cilla G, Montes M, Gomariz M, Alkorta M, Iturzaeta A, Perez-Yarza EG, et al. Rotavirus genotypes in children in the Basque Country (north of Spain): rapid and intense emergence of the G12[P8] genotype. *Epidemiol Infect* 2013;141(4):868–874.
- Luchs A, Cilli A, Morillo SG, Carmona R, Timenetsky M. Rotavirus genotypes circulating in Brazil, 2007–2012: implications for the vaccine program. *Rev Inst Med Trop* Sao Paulo 2015;57(4):305–313.
- Neves MA, Pinheiro HH, Silva RS, Linhares AC, Silva LD, Gabbay YB, et al. High prevalence of G12P[8] rotavirus strains in Rio Branco, Acre, Western Amazon, in the post-rotavirus vaccine introduction period. J Med Virol 2016;88(5):782–789.
- Tort LF, Victoria M, Lizasoain AA, Castells M, Maya L, Gomez MM, et al. Molecular epidemiology of group A rotavirus among children admitted to hospital in Salto, Uruguay, 2011–2012: first detection of the emerging genotype G12. J Med Virol 2015;87(5):754–763.

- 35. Delogu R, Ianiro G, Camilloni B, Fiore L, Ruggeri FM. Unexpected spreading of G12P[8] rotavirus strains among young children in a small area of central Italy. *J Med Virol* 2015;87(8):1292–1302.
- 36. Ndze VN, Esona MD, Achidi EA, Gonsu KH, Doro R, Marton S, et al. Full genome characterization of human Rotavirus A strains isolated in Cameroon, 2010–2011: diverse combinations of the G and P genes and lack of reassortment of the backbone genes. *Infect Genet Evol* 2014;28:537–560.
- Mijatovic-Rustempasic S, Teel EN, Kerin TK, Hull JJ, Roy S, Weinberg GA, et al. Genetic analysis of G12P[8] rotaviruses detected in the largest U.S. G12 genotype outbreak on record. *Infect Genet Evol* 2014;21:214–219.
- Aly M, Al Khairy A, Al Johani S, Balkhy H. Unusual rotavirus genotypes among children with acute diarrhea in Saudi Arabia. BMC Infect Dis 2015;15:192.
- Tiku VR, Sharma S, Verma A, Kumar P, Raghavendhar S, Aneja S, et al. Rotavirus diversity among diarrheal children in Delhi, India during 2007–2012. Vaccine 2014;32 Suppl 1:A62–A67.
- Kirkwood CD, Boniface K, Barnes GL, Bishop RF. Distribution of rotavirus genotypes after introduction of rotavirus vaccines, Rotarix(R) and RotaTeq(R), into the National Immunization Program of Australia. Pediatr Infect Dis J 2011;30(1 Suppl):S48–S53.
- Leshem E, Lopman B, Glass R, Gentsch J, Bányai K, Parashar U, et al. Distribution of rotavirus strains and strain-specific effectiveness of the rotavirus vaccine after its introduction: a systematic review and meta-analysis. Lancet Infect Dis 2014;14(9):847–856.
- Zeller M, Patton JT, Heylen E, De Coster S, Ciarlet M, Van Ranst M, et al. Genetic analyses reveal differences in the VP7 and VP4 antigenic epitopes between human rotaviruses circulating in Belgium and rotaviruses in Rotarix and RotaTeq. J Clin Microbiol 2012;50(3):966– 976.
- 43. Matthijnssens J, Zeller M, Heylen E, De Coster S, Vercauteren J, Braeckman T, et al. Higher proportion of G2P[4] rotaviruses in vaccinated hospitalized cases compared with unvaccinated hospitalized cases, despite high vaccine effectiveness against heterotypic G2P[4] rotaviruses. Clin Microbiol Infect 2014;20(10):O702– O710.
- 44. Pitzer VE, Bilcke J, Heylen E, Crawford FW, Callens M, De Smet F, et al. Did large-scale vaccination drive changes in the circulating rotavirus population in Belgium? Sci Rep 2015;5:18585.
- 45. Donato CM, Cowley D, Donker NC, Bogdanovic-Sakran N, Snelling TL, Kirkwood CD. Characterization of G2P[4] rotavirus strains causing outbreaks of gastroenteritis in the Northern Territory, Australia, in 1999, 2004 and 2009. Infect Genet Evol 2014;28:434–445.